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APOE at the interface of inflammation, neurodegeneration and pathological protein spread in Alzheimer's disease

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Keywords: Apolipoprotein E; APOE; Alzheimer's disease; inflammation; glia; tau; neurodegeneration

Abbreviations

Alzheimer's disease (AD); Amyloid beta (A β); Apolipoprotein E (APOE); Disease Associated Microglia (DAM); Glial fibrillary acidic protein (GFAP); Human Leukocyte Antigen-DR (HLA-DR); Interleukin 1 β (IL-1 β); Interleukin 6 (IL-6); Lipopolysaccharide (LPS); Macrophage Scavenger Receptor-A (MSR-A); Oligomeric amyloid beta (oA β); Prostaglandin E2 (PGE2); Triggering Receptor Expressed on Myeloid cells 2 (TREM2); Tumour Necrosis Factor α (TNF- α)

Short running title

Review of APOE in Alzheimer's disease pathogenesis

Abstract

Despite more than a century of research, the aetiology of sporadic Alzheimer's disease (AD) remains unclear and finding disease modifying treatments for AD presents one of the biggest medical challenges of our time. AD pathology is characterised by deposits of aggregated amyloid beta (A β) in amyloid plaques and aggregated tau in neurofibrillary tangles. These aggregates begin in distinct brain regions and spread throughout the brain in stereotypical patterns. Neurodegeneration, comprising loss of synapses and neurons, occurs in brain regions with high tangle pathology, and an inflammatory response of glial cells appears in brain regions with pathological aggregates. Inheriting an apolipoprotein E ϵ 4 (*APOE4*) allele strongly increases the risk of developing AD for reasons that are not yet entirely clear. Substantial amounts of evidence support a role for APOE in modulating the aggregation and clearance of A β , and data have been accumulating recently implicating APOE4 in exacerbating neurodegeneration, tau pathology and inflammation. We hypothesise that APOE4 influences all the pathological hallmarks of AD and may sit at the interface between neurodegeneration, inflammation and the spread of pathologies through the brain. Here we conducted a systematic search of the literature and review evidence supporting a role for APOE4 in neurodegeneration and inflammation. While there is no direct evidence yet for APOE4 influencing the spread of pathology, we postulate that this may be found in future based on the literature reviewed here. In conclusion, this review highlights the importance of understanding the role of APOE in multiple important pathological mechanisms in AD.

Introduction

The greatest genetic risk factor for sporadic AD is a polymorphism in the apolipoprotein E (*APOE*) gene. The *APOE* ϵ 4 (*APOE4*) allele is associated with increasing risk of AD in a dose-dependent manner when compared to the more common *APOE* ϵ 3 (*APOE3*) allele; whereas the much rarer *APOE* ϵ 2 (*APOE2*) allele has been shown to be protective [1]. The inheritance of two copies of *APOE4* increases the chance of developing AD by 12 times compared to the risk of a person with two copies of *APOE3*. Homozygous *APOE4* carriers who develop AD also have a lower average age of clinical onset of 68 years of age compared to an average age of onset of 84 for an individual with two copies of *APOE3*. One copy of *APOE4* increases the chance of AD by 3 times and lowers the average age of onset to 76 years of age [1]. Although mentioned in association with AD most frequently, APOE has also been linked to Parkinson's Disease [2], Frontotemporal dementia [3] and other neurological diseases (reviewed in [4]) as well as linked to lower cognition in non-demented aged individuals [5]. The pathways by which APOE impacts the development of AD have been widely studied both *in vitro* and *in vivo* however the exact mechanisms have yet to be uncovered.

Much of the work looking at APOE in AD investigates its relationship with A β . Early post mortem work found a positive correlation between *APOE4* allele dose and A β plaque density in individuals with AD [6]. A wide range of compelling studies indicate that APOE4 affects the production, clearance and aggregation of A β (reviewed in [4,7]). Recent genetic data that strongly suggest inflammation to play a role in AD risk have re-invigorated the investigations of the role of APOE in neuroinflammation and how this contributes to disease [8]; and there are emerging data suggesting that APOE may also influence tau-mediated neurodegeneration [9].

We hypothesise that APOE4 acts beyond its well-known roles in influencing A β pathology and lipid homeostasis and has a strong influence on neurodegeneration, inflammation and potentially the spread of pathological proteins through the brain. Here we conducted a systematic literature search and review the current support in the literature for this hypothesis.

Methods

A systematic literature search approach was taken for finding studies to review in this paper. In February 2018, Embase, Web of Science and MedLine were searched to identify primary research articles published from 1980 to the date searches were run. Search terms covering APOE, AD and inflammation/pathological protein spread/neurodegeneration were developed (Supplemental Table 1) to suit each database. Initially, there were no language or selection restrictions on the type of study included or how outcomes were defined, measured or when they were taken. Searches identified 22,909 abstracts and titles that were exported to Endnote, where 12,638 duplicates were removed. 10,271 articles were uploaded into Covidence, where a further 767 duplicates were removed. A two-stage screening strategy was conducted on titles/abstracts and then on full-texts using pre-determined exclusion criteria (Supplemental Table 2). Three researchers contributed to the abstract and title screening process such that 50% of abstracts/titles were screened by at least 2 people and 50% by one. All full-text articles were double screened. Of the 9502 titles/abstracts screened, 214 progressed to full-text review and 88 studies were included. 20 hand-picked papers that were either published after the search date or were missed during the search but deemed pertinent to the review were also included as is standard practice for full systematic reviews (Figure 1; Supplemental Table 3). In the results section, we synthesize the findings of the papers identified by the systematic search. Due to the heterogeneous nature of the studies, we did not perform standardised quality control checks of all of the papers, however all papers included were published in peer-reviewed journals. Because systematic reviews are designed to test evidence of an intervention and due to the inability to conduct formal quality control due to the many types of experiments reviewed, this is not a fully registered systematic review but instead uses some of the principles of systematic reviews to perform a *systematic literature search* and screen for relevant papers which we review.

Results of the systematic literature search

APOE and Neurodegeneration

In order to determine whether there is evidence that APOE influences neurodegeneration search terms were used to identify papers containing both APOE and indicators of neuron and synapse loss (papers relating to APOE and neurodegeneration identified in the systematic search are coloured blue and orange in Supplemental Table 1).

Imaging studies in AD populations repeatedly demonstrated possession of an *APOE4* allele to be associated with more extensive atrophy in disease-specific brain regions such as the medial temporal lobe [3,11–23], although this was not universal [24,25]. This is unsurprising when considering that brain structures located here have clinical correlates to well-defined AD symptoms (e.g. memory impairments – hippocampus; altered emotional responses - amygdala), and that *APOE4* carriers are at increased risk of developing these symptoms in the form of AD. *APOE4*-related atrophy of medial temporal lobe structures was suggested to occur in a gene-dose-dependent manner [15–17,19,21], with one study finding each *APOE4* allele to impart a 4.8% reduction in hippocampal volume and a 3.8% reduction in amygdala volume [16]. In longitudinal studies, accelerated rates of hippocampal atrophy were associated with the *APOE4* allele [13,22,26–29], although this was not always observed [30,31]. *APOE4*-related atrophy was also observed in the parietal cortex [3,13] and some prefrontal areas [3,12], although again these were not consistently reported and may be related to the inclusion of younger AD patients in these studies.

Diffuse cerebral atrophy is a gross pathological feature of AD. Somewhat surprisingly, however, whole brain volume has been proposed to increase with increasing number of *APOE4* alleles in AD patients [19,32]. Greater *APOE4* allele dose was associated with larger volumes in the frontal lobes [11,17,20], although not always [12,14], which might potentially outweigh reductions in other brain regions and account for increased whole brain volume. Nonetheless, these studies indicate that the *APOE4* allele selectively influences the topography of regional brain atrophy, and thus neurodegeneration, in AD.

Studies of post-mortem human AD brain have identified neuronal loss in vulnerable brain regions to be characteristic of AD. From our literature search, determining whether APOE influences this was less clear. In the nucleus basalis, *APOE4* was found to exacerbate neuronal loss [33], but have no impact in other studies [34,35]. These discrepancies may be due to the different methods employed in quantitation of neuronal loss and groups studied. *APOE4* allele possession was also found to enhance neuronal loss in other subcortical structures [33,34], but have no effect on the CA1 region of the hippocampus or superior temporal sulcus [34,36].

In vivo and *in vitro* studies supported an association between APOE4 and neuronal degeneration. In aged mice expressing human APOE, those expressing APOE4 exhibited increased hippocampal and cortical atrophy compared to those expressing APOE3 [37,38]. In APOE4, but not APOE3 mice, activation of the amyloid cascade by inhibition of the A β degrading enzyme neprilysin was sufficient to induce degeneration of hippocampal and entorhinal cortex neurons, suggestive of a specific effect of APOE4 in exacerbating A β -related neuronal loss [39]. Isoform-specific effects of APOE on A β -induced neurodegeneration were also suggested *in vitro*, with APOE3 and APOE2, but not APOE4, protecting hippocampal and cortical neurons from A β -induced neurotoxicity [40–42]. APOE4 has also been shown to exacerbate neurodegeneration in the absence of A β , instead operating through a tau-dependent mechanism [9].

Animal studies support an isoform-specific role for APOE in the loss of GABAergic interneurons [43,44]. Mice expressing human APOE4 exhibited greater age-dependent loss of GAD67- and somatostatin-positive interneurons in the dentate gyrus compared to those expressing APOE3. Interestingly, loss of somatostatin immunoreactivity was exacerbated by APOE4 in the AD brain [45]. The detrimental effects of APOE4 on GABAergic interneurons was sex-dependent, only being observed in female mice [44]. This is interesting when considering APOE4 confers greater AD risk in females and some effects of APOE4 on regional brain atrophy are more prominent in females [16,18,21].

In summary, there is strong evidence that APOE4 influences neuron loss in AD, however there are some conflicting reports and future well-powered, rigorous studies in both human brain and animal models are needed to fully understand the age, sex, and region-specific effects of APOE on neuron death.

APOE-related synaptic and dendritic degeneration in AD

Prior to the onset of neuronal loss in AD, extensive synapse loss and dendritic changes occur. These can be considered early neurodegenerative processes that contribute to synaptic and neuronal dysfunction, which pave the way for more generalised neurodegeneration later in the disease. In contrast to neuronal loss, synaptic and dendritic degeneration are dynamic processes, with the potential to be reversed if targeted early enough. Thus, it is important to consider the role of APOE in these changes.

Whether synapse loss associates with *APOE* genotype was the subject of a few studies identified by our search. Electron microscopy coupled with stereological counting in post-mortem tissue identified AD-related loss of synapses in the dentate gyrus, stratum radiatum of the CA1 region of the hippocampus and lamina III of the inferior temporal gyrus. However, this was not related to *APOE* status [46–48]. These findings contrast with results from aged *APOE* mice, which suggested *APOE4* to be associated with a reduced number of synapses in the dentate gyrus [49]. Although not accounting for these discrepancies, it is worth noting that electron microscopy is limited by its ability to only quantify synaptic changes in small areas of tissue. Thus, its use for characterising changes in a disease that stereotypically results in widespread pathology may not be entirely representative.

In AD, loss of the presynaptic vesicle protein, synaptophysin, is evident in various brain regions. In post-mortem studies, *APOE* genotype did not modulate synaptophysin levels in frontal or temporal regions [50,51] although a trend towards lower synaptophysin immunoreactivity was observed in AD patients with an *APOE4* allele [50]. A further study of another presynaptic vesicle protein, Rab3a, also found no association of synapse loss with *APOE* genotype [52]. This contrasts with animal studies demonstrating that mice expressing human *APOE4* alone or in concurrence with human amyloid precursor protein display increased age-dependent degeneration of synaptophysin-positive presynaptic terminals in the neocortex and hippocampus [53,54]. Preservation of synaptophysin-positive presynaptic terminals in aged *APOE4* mice has also been reported, however [55]. Contradictory results have also been reported in *APOE* KO mice, with both age-dependent reductions in synaptophysin-positive terminals [56] and no changes [57] observed.

Although the findings discussed thus far tend to suggest that *APOE4* does not contribute to synaptic degeneration, it is important to consider that techniques used may not be optimal for quantification of synapses or synaptic protein loss, and that many of these studies did not consider plaque proximity in the analyses which is known to drive substantial local synapse loss as will be discussed below. The axial resolution of even confocal microscopy is not sufficient to resolve individual synapses in standard tissue sections. In addition, immunoblotting is inferior to techniques that yield absolute synaptic protein concentrations. Indeed, using ELISA, pre-synaptic protein levels were found to be reduced in AD patients with an *APOE4* allele, although only a trend towards reduction was seen for synaptophysin [58].

A relatively recently described histological technique, array tomography, has been used to overcome some of the limitations associated with synapse quantification using other methods, offering a means for high-resolution characterisation of synapses in post-mortem tissue. Additionally, this approach avoids some issues associated with electron microscopy in that thousands of synapses can be analysed. Using this technique, synapse density was found to be specifically reduced within the 'halo' surrounding amyloid plaques. In both AD human post-mortem tissue and a mouse model, this was isoform-specific, with APOE4 exacerbating peri-plaque synapse loss [59,60].

APOE4-related peri-plaque synapse loss affects both pre- and post-synapses [59] and greater age-induced reductions in post-synaptic proteins have been observed in AD mouse models expressing APOE4 compared to other isoforms [61]. Dendritic abnormalities, such as dystrophic neurites, alterations in dendrite complexity and loss of spines are widespread in AD. Considering that dendritic abnormalities are closely linked to synaptic dysfunction, and thus potentially synaptic degeneration, the effect of APOE here is relevant.

The presence of dystrophic neurites are a neuropathological hallmark of AD and are exacerbated by APOE4 in AD mouse models when compared to other isoforms [59]. Such changes in dendritic morphology are predicted to alter synaptic function, and thus may influence degeneration. Indeed, neuritic degeneration requires the presence of APOE [62], with APOE4 mice showing increased age-dependent loss of neocortical and hippocampal dendrites compared to APOE3 mice [53].

The density of dendritic spines, the post-synaptic site of over 90% of excitatory neurons, has consistently been shown to be reduced in the presence of APOE4 in cortex, hippocampus, entorhinal cortex and amygdala *in vitro* [63], *in vivo* [64–68] and in humans [68]. A reduction in dendrite length was also observed in APOE4 mice [66,67], which contributes to reduced connectivity. Interestingly, expression of APOE2 can rescue reduced spine density in AD mouse models [69]. The morphology of dendritic spines has also been suggested to be influenced by APOE, with APOE4 being associated with shorter spines and APOE2 with longer spines [64]. Additionally, APOE may influence spine morphology such that APOE4 specifically reduces the number of spines associated with learning and memory [65]. Finally, APOE4 is associated with reductions in dendritic arborisation and less complicated branching patterns, impacting on neuronal function [64,65,67].

Collectively, these studies suggest that, in the context of dendritic and synaptic changes, APOE4 is less effective at maintaining synaptic and neuronal integrity in disease-specific brain areas than other isoforms, which likely contributes to synaptic and neuronal degeneration.

Mechanisms of APOE-related neurodegeneration

Dendritic, synaptic and neuronal degeneration are all influenced by APOE in an isoform-specific manner. However, the mechanisms by which APOE4 impacts neurodegeneration are not completely understood. Elucidating these mechanisms may enable the development of appropriate therapies, particularly in relation to APOE effects on synapses and dendrites, which have potential to be reversible.

In vivo and *in vitro* studies suggested that APOE4 may drive neurodegeneration through an A β -dependent mechanism [37,39–42]. Recent revisions of the amyloid hypothesis of AD, have suggested soluble oligomeric forms of A β (oA β) to be the effectors of A β -induced degeneration, with APOE4 exacerbating oA β -associated degeneration relative to other isoforms [41,42,70] (Figure 2) (details of participants in Figures 2-5 found in Supplementary Table 4). APOE and oA β may act intracellularly to enact this degeneration, with APOE uptake into neurons correlating with neuronal death and intracellular accumulation of soluble A β [71]. APOE4-specific increases in intraneuronal oA β have been suggested to drive neurodegeneration through impairments of mitochondria and lysosomes [39,72], although the role of intracellular A β remains hotly debated in the field. Isoform-specific interactions between APOE and the C-terminal domain of soluble A β , or lack of in the case of APOE4, have also been suggested to influence the propensity for APOE4 to promote A β -mediated neuronal death [42]. Moreover, protection against oA β -mediated synaptic loss by APOE3 has been suggested to be mediated by a novel intracellular protein kinase C pathway, which is not activated by APOE4 [70].

APOE and oA β may also act extracellularly to impact neurodegeneration. Interestingly, oligomeric forms of A β are increased in the ‘halo’ surrounding plaques, an area where synaptic degeneration is exacerbated by APOE4 [59,60]. Mechanistically, APOE4 might exacerbate peri-plaque synapse loss by facilitating the association of oA β with synapses where it is toxic, thus resulting in synapse loss [60].

While APOE certainly influences A β -dependent neurodegeneration, it is becoming increasingly clear that there are other mechanisms by which APOE influences degeneration (Figure 3). For example, in mice expressing a form of mutant human tau associated with frontotemporal dementia, co-expression of human APOE4 led to a drastic increase in neurodegeneration compared to that seen with other APOE isoforms. APOE was necessary for this tau-mediated neuronal death to occur [9], suggestive of APOE exerting a neurodegenerative effect through tau. APOE4 has also been shown to impair GABAergic interneurons through a tau-dependent mechanism *in vivo* [43]. Although known to be related to A β , dysregulation of calcium homeostasis has also been implicated as a mechanism of APOE-related neurodegeneration, independent of A β . Here, APOE4 increased levels of cytosolic calcium, an effect that was dose-dependently associated with cell death [73].

Another important consideration, and one that is often neglected, is the cellular source of APOE. Although primarily produced by glial cells, under conditions of stress or injury neurons also synthesise APOE [74]. Considering that the AD brain can be considered both a 'stressed' and 'injured' environment, the cellular source of APOE is likely to be relevant to its effect on neurodegeneration. Indeed, at the level of the dendrite, loss of spines, reduced arborisation and alterations in morphology were observed in mice expressing neuronal APOE4, but not astrocytic APOE4 [65,75]. This phenomenon has also been confirmed to occur at the level of the synapse, with only neuronal APOE4 promoting the degeneration of presynaptic terminals and cell death [75]. Consequently, the cellular source of APOE seems to impact upon its capacity to induce neurodegeneration. Further studies are needed to examine how neuronal and astrocytic APOE differ from one another, to characterise how these effects are mediated.

A potential mechanism by which neuronal APOE4 exerts increased neurotoxic effects may be due to the intraneuronal proteolytic processing of APOE, whereby APOE can be cleaved to generate C-terminally-truncated fragments. In the human AD brain, these fragments are more numerous in individuals carrying an *APOE4* allele. *In vitro*, APOE4 is more susceptible to proteolytic cleavage than APOE3 and, in mice, these APOE fragments are capable of eliciting AD-like neurodegeneration [76]. Further work has identified the lipid-binding region of APOE to be essential for toxicity although not sufficient alone. Instead, the lipid- and receptor-binding regions appear to act in concert to mediate toxic effects [77]. APOE4 fragments have also been shown to interact synergistically with A β and tau,

exacerbating both pathologies and increasing the degree of neurodegeneration [43,78]. In relation to A β , APOE4 fragments bind poorly to A β leading to reduced clearance and increased deposition [78] and these fragments also promote intraneuronal A β accumulation [79]. In relation to tau, APOE fragments increase the degree of phosphorylation, likely exacerbating neurodegeneration [43]. Cleavage fragments of APOE may also exert neurodegenerative effects by increasing intracellular calcium [80], a mechanism that has previously been associated with full-length APOE4, as aforementioned, or by impairing mitochondrial function and integrity [77]. Finally, structural differences among APOE isoforms may also contribute to the more neurodegenerative phenotype associated with APOE4. Unlike APOE2 and APOE3, APOE4 exhibits a domain interaction, whereby a salt bridge mediates interaction between N- and C-terminal domains. Indeed, in mice, this domain interaction has been shown to be associated with pre- and post-synaptic protein loss [81], thus suggesting another mechanism by which APOE4 might contribute to synaptic pathology and neurodegeneration.

APOE genotype may further contribute to synaptic degeneration, and subsequent neuronal degeneration, through its effects on dendrites. Dendritic changes are suggested to alter neuronal plasticity and regenerative capacity, leading to synaptic dysfunction and subsequent synapse and neuron loss. Indeed, more plastic changes are seen in the human AD brain in the absence of an *APOE4* allele [33]. Various mechanisms have been proposed as to how APOE isoforms differentially regulate dendritic changes in AD. These range from altered binding of APOE to receptors and subsequent intracellular signalling cascades [82,83], impaired regulation of receptors within spines [63], elevations in calcineurin activity that is associated with reductions in spine density [84] and impairments of neuronal outgrowth [82], among others.

Overall, these results highlight the far-reaching and diverse biological effect APOE4 has on neurodegeneration in AD with particularly strong evidence supporting a role for APOE4 in synapse degeneration.

APOE and Inflammation

In order to determine whether there is evidence that APOE influences inflammation, search terms were used to identify papers containing both APOE and indicators of inflammation (papers from the systematic search that support a role for APOE in inflammation are coloured in green and orange in Supplemental Table 1).

Increased glial activation and gliosis with APOE4

There is substantial reactive glial cell accumulation, termed gliosis, during AD which is enhanced in the presence of the *APOE4* allele (Figures 4 and 5). Markers of glial activation are commonly used to reflect a variety of functional outcomes and to quantify the changes in glial numbers and their respective phenotype.

Human post-mortem studies have quantified gliosis in different brain regions using multiple markers of activation. By immunophenotyping microglia in the frontal gyrus and correlating to APOE status in AD and control cases, markers of activation (CD68, Human Leukocyte Antigen-DR isotype [HLA-DR], CD64) were found to be significantly associated with APOE4 carriers while APOE2 carriers were associated with higher levels of more homeostatic microglial markers (Iba1 and Macrophage Scavenger Receptor-A; MSR-A) and lower levels of the reactive ones [85]. Although the microglial phenotype was insufficient in predicting the APOE status in dementia, the elevated markers of phagocytosis, adaptive immune response and antigen recognition seen in APOE4 carriers point towards a more pathological environment. Other brain regions have been assessed for microglial levels, where in an *APOE4* dose-dependent manner, there was extensive microgliosis in both the frontal and temporal cortices [86]. Similarly, there is elevated GFAP-positive astrogliosis in the grey matter of APOE4 AD patients compared to other *APOE* genotypes, and an overall greater astrogliosis between AD and age-matched control cases [87]. Interestingly, the *APOE4* allele was associated with differences in GFAP burden in non-demented individuals, despite the increased plaque burden of non-demented aged APOE4 individuals, supporting the importance of glial cells in AD pathogenesis. As always, the post-mortem tissue only provides a snap-shot of end-stage of disease, so we rely on mouse models for deciphering the mechanistic changes that APOE4 induces during AD.

Mouse studies looking at the effects of the human *APOE* allele with AD-like pathology have mostly replicated the gliosis data seen in human post-mortem tissue. APOE4 mice crossed to the 5xFAD amyloidosis model showed increased microgliosis in deep cortical layers accompanied by a greater number of dystrophic microglial processes in the presence of the *APOE4* allele, compared to *APOE4* and *APOE3* [88]. Of note, both APOE2 and APOE4 5xFAD mice had more A β -associated microglia than APOE3 [88,89], suggesting that the APOE2 conformation may be protective not by preventing microglia/plaque interactions but by mediating more effective ways to respond to the plaques and

making microglia more resistant to amyloid toxicity. These changes were not observed in the subiculum of these mice, reiterating regional differences in microglia. Furthermore, when APOE knock-in mice were crossed to a tauopathy model (P301S), CD68-positive microglial burdens in the hippocampus and entorhinal/piriform cortex were markedly increased in APOE4 mice compared APOE3 and APOE knock-out mice in a tau pathology mediated manner [9]. The same effect was also observed with GFAP immunoreactivity, further establishing an aberrant glial response to AD-like pathology in combination to the presence of the *APOE4* allele. Similarly, there was neurodegeneration-associated microgliosis and astrogliosis in the hippocampus of APOE4 mice, compared to APOE3, but no differences were found in the septum [90]. However, in the hippocampus of APOE4 LPS-injected mice there was marked microgliosis although astrogliosis was found in the APOE3 mice [91]. In summary, in APOE4 amyloidosis (5xFAD) and tauopathy (P301S) mouse models there was exacerbated gliosis. Although, there was important regional variability, taken together these data strongly support a role for APOE4 in promoting inflammatory changes in microglia and astrocytes.

The precise mechanisms by which reactive gliosis is established in the APOE4 AD brain is unknown, but a key question that remains is whether this gliosis is a driver of the disease, accounting for the earlier onset and worse prognosis *APOE4* carriers face, or a by-product of the exacerbated amyloid and tau pathology. To answer these questions, the inflammatory capacity of glial cells and their transcriptomic signatures in the presence of the APOE4 are being currently assessed.

ApoE-related glial transcriptional changes

Transcriptional studies are becoming increasingly popular in the microglia and neuroinflammation field, with *ApoE* upregulation consistently being a top hit in AD-like mouse models. The advantages of RNA-sequencing are a non-biased, high-yield output of all transcriptional changes as well as the ability to use these long data sets to investigate biological pathways. As such, this process speeds up the identification of genetic and molecular pathways involved in AD pathogenesis and ways they can be therapeutically targeted.

Recently, microglial *Apoe* mRNA transcript levels were quantified in two models of AD-like pathology (amyloidosis and tauopathy) and ageing [92]. Its high abundance in all conditions supports a role for APOE as a key part of the microglial signature, despite its expression not being restricted to microglial cells. In terms of its relative expression, *Apoe* was highly upregulated in aging and disease models, with aging female mice showing a marked increase. Pathway analysis puts APOE as the driver of a network whose downstream effectors are also highly upregulated in these models, like the chemoattractant CCL3, whose relevance to neuroinflammation will be discussed in the next section if this review.

Apoe transcription is also downstream of the activation of a microglial receptor TREM2 (Triggering Receptor Expressed on Myeloid cells 2), another AD risk gene [93]. This APOE activation pathway results in a more pro-inflammatory microglial response and a degenerative phenotype, as seen in the AD brain. Microglial and astrocytic pro-inflammatory genes were also profoundly upregulated in APOE4 KI mice crossed with a tauopathy model (P301S), compared to APOE3 [9]. Conversely, APOE knock-out/P301S mice showed attenuation of this impaired pro-inflammatory profile, highlighting APOE as a master regulator of glial inflammatory response with its *APOE4* allele being associated to a pro-degeneration phenotype.

Single-cell RNA sequencing takes this a step further by identifying and characterising clusters of subpopulations within a cell type, for instance disease-associated microglia (DAMs) [94]. Specifically, *Apoe* is upregulated early in DAMs of the 5xFAD AD-like model, even in the absence of TREM2. A TREM2 independent pathway is thus proposed to initiate *Apoe* upregulation in the early phase of AD, with a later TREM2-dependent pathway activating *Apoe* transcription which induces neurodegenerative microglia. This potentially forms a therapeutic window where preventing the second *Apoe* induction via TREM2 may protect against the exacerbated inflammation and degeneration caused by microglia. Understanding the ways in which TREM2 is activated in the AD brain and the effectors mediating this TREM2-APOE pathway can provide new ways to halt AD progression and hinder neuroinflammation.

Human induced pluripotent stem cells (iPSCs) derived from AD patients were recently transcriptionally characterised after directing them towards a microglial-like lineage [95]. Importantly, the microglia-like cells were engineered with Crispr-Cas9 to correct APOE4 into APOE3.

Not only did this result in attenuation of AD-associated morphological and transcriptomic signatures, but immune-related genes were also upregulated.

Overall, *ApoE* is consistently one of the most upregulated genes in transcriptomic studies of microglia in AD-like models. Nevertheless, a limitation of transcriptomics involves the isolation process which pushes microglia into an activated phenotype [92]. There is a lack of detailed studies of human post-mortem tissue profiling transcription in cases with different APOE genotypes, which will be needed to confirm the translational relevance of mouse studies. As this is still at the transcriptional level, studies looking at a more functional level are imperative to understand how these RNA changes relate to disease.

Altered inflammatory response by glial cytokine release

Innate and adaptive immune cells respond to environmental stimuli by releasing signalling cytokines and chemokines. Our systematic literature search showed that the process of cytokine release by glia to maintain homeostasis and respond to damage is dysfunctional in the aging and AD brain [96], with substantial evidence pointing towards the *APOE4* allele playing a crucial role in this [97–99], supporting the transcriptional profile changes seen in $\epsilon 4$ microglia and astrocytes.

APOE4 alters the baseline pro-inflammatory response even in the absence of disease. Addition of APOE4, but not APOE3 protein, to rat microglia cultures alone stimulated the secretion of prostaglandin E2 (PGE2), interleukin-1 β (IL-1 β), and nitric oxide (NO) [100–103]. On the other hand, microglial and astrocyte stimulation with APOE4 and A β attenuated the production of inflammatory mediators [103–105], indicating a more complex interaction of microglia, APOE, and A β *in vivo*. A physiological concentration of A β may therefore be beneficial for glial functioning by interacting with APOE.

LPS-activated microglia and astrocytes are a well-characterised model of glial activation by mimicking the inflammation seen in AD. Microglia induced with LPS in the APOE4 background released greater amounts of pro-inflammatory cytokines, like tumour necrosis factor- α (TNF- α), IL-1 β , and interleukin-6 (IL-6) [106], an effect replicated in AD-like models [88,107]. Simultaneously,

APOE4 treated LPS-induced microglia suppressed the production of TNF- α less than the APOE3 and APOE2 isoforms [108] while APOE-/- mice secrete lower levels of anti-inflammatory cytokines [109], showing a physiological role of APOE in modulating inflammation. Indeed, knocking out murine *ApoE* increases glial production of nitric oxide (NO) [110] and other inflammatory mediators, like *TNF- α* , *IL-1 β* and *IL-6* transcripts in the CNS [111]. Murine *ApoE* is therefore required to suppress glial-mediated inflammation providing a physiological role in CNS homeostasis, which is disrupted in AD, potentially in an age-dependent manner. This evidence points to APOE4 expressing microglia being both more pro-inflammatory and anti-inflammatory at the same time.

In contrast to the microglial data, astrocytes from APOE2 and APOE3 animals treated with LPS produced more of these pro-inflammatory cytokines than the APOE4 counterparts (IL-1 β , TNF- α , and IL-6) [112]. APOE4 astrocytes also produce more CCL3 (chemokine C-C motif ligand 3) [113], similar to microglial *Ccl3* mRNA upregulation in AD-like models [92], which is downstream of the APOE-driven network while APOE-/- mice produce less CCL3 [109]. Despite the increase of CCL3, the chemoattraction ability of microglia is impaired in the presence of the *APOE4* allele, as they are less migratory and receptive to immune sensing [114–116]. Data so far suggest that APOE4 confers a more pro-inflammatory and less anti-inflammatory phenotype in microglia, with an opposing pattern in astrocytes.

APOE at the interface of inflammation and neurodegeneration: glial-mediated synapse loss

The synaptic loss during the early phase of AD is now thought to be partly due to aberrant microglial and astrocyte complement-mediated phagocytosis [117,118]. Given APOE's role in synapse loss and inflammation, we postulate that the APOE4 genotype is implicated in synaptic loss through a glial-mediated mechanism (Figure 6).

LPS intracerebral injections in APOE4 mice led to decreased pre- and post-synaptic protein levels as well hippocampal gliosis and pro-inflammatory cytokine release [119]. Moreover, APOE4 is accompanied by greater complement activation [120], which is the proposed synaptic tag for synaptic clearance. Still, this evidence is correlative and there are other mediators than could affect synaptic loss. In development, astrocytes of the APOE4 background were less phagocytic towards

pHrodo-labelled synaptosomes than those of APOE3 and APOE2 [121], leading to the hypothesis that some synapses are not pruned by APOE4 astrocytes, accumulating complement and making them more vulnerable in AD. Whether these synapses are defective or not is a key question, as loss of healthy synapses with accumulated complement would explain the initial synapse loss in AD, and the earlier onset of APOE4 carriers.

Although a lot more evidence is required, particularly from the human perspective, to understand if and how glial cells drive the synapse loss during AD, understanding why APOE e4 carriers at greatest risk have this extensive synapse loss and greater onset will be crucial to tailor therapies for individuals of this genotype.

Potential role for spread of pathological proteins through the brain

No studies were found in the systematic search that specifically investigated the role of APOE in pathological protein spread. Two of the current hypotheses about the spread of tau through the brain are that tau spreads trans-synaptically and that microglia eat tau-containing synapses facilitating its spread (Figure 7). Interestingly, in this review APOE genotype is shown to affect both tau and microglia in multiple ways, implicating APOE4 as a potential facilitator of misfolded proteins spreading between brain regions.

Conclusion

Our systematic literature search revealed strong links between APOE and synapse degeneration, which is further supported by relevant literature that did not fall into our search terms. The impaired phagocytic capacity of TREM2^{-/-} microglia for synapses and synaptosomes in development [122] highlights yet again the TREM2/APOE pathway as a potential AD-related mechanism of synaptic elimination. The interplay between glia cell types in relation to APOE genotype is also a new avenue to be assessed considering that astrocytic IL-33 induces microglial synaptic engulfment of both excitatory and inhibitory synapses [123]. Conversely, microglia have been shown to induce a neurotoxic astrocyte phenotype via IL-1 α , C1q, and TNF [124], the two latter being increased with

APOE4 expression. Moreover, APOE4-elevated soluble factors released by microglia like NO and IL-6 can induce synapse loss in neuron culture systems [125], suggesting microglia may alter synaptic numbers both by phagocytosis and their secretome.

In studies identified by our systematic search, APOE4 was consistently associated with increased neurodegeneration within the medial temporal lobe; this region includes the entorhinal cortex, where tau pathology begins [126]. APOE4-associated degenerating synapses may release tau seeds via exosomes, stress-produced nanotubes [127], or passively as terminals degenerate. These may be taken up by recipient cells therefore propagating tau trans-synaptically. However, our recent data indicate that presynaptic terminal degeneration is not necessary for the spread of tau through neural circuits [128], making it important to investigate other mechanisms of pathological protein spread.

Microglia have also been suggested to mediate tau spread [129]. The exacerbated neurodegeneration in APOE4 likely induces microglia and astrocytes to phagocytose degenerating tau-containing synapses and neurons. Exosomes synthesised from these microglia, containing synaptic-impairing micro-RNA [130] and tau seeds, may propagate tau pathology and neurodegeneration [131]. Astrocytes may also be involved in this, given their role in α -synuclein transfer via tunnelling nanotubes in Parkinson's disease [132].

APOE, therefore, is potentially at the interface of inflammation, neurodegeneration, and the pathological protein spread (summarized in figures 6 and 7). Tau pathology and synapse loss are the strongest correlates with cognitive impairments [133]; therefore, preventing these processes could have significant impact on disease progression. Further evidence is, however, needed to directly link APOE to these spreading mechanisms.

Our systematic literature search and review of the resulting papers highlights the need for multiple approaches to understand the complex role of APOE in disease, as has been observed recently for many fields in science [134]. Many findings remain contradictory and will need further support and investigation using different model systems to warrant moving forward to therapies targeting APOE. These caveats notwithstanding, our review of the literature supports the idea that understanding

how APOE influences multiple pathological features of AD will be important for developing effective therapeutics to prevent or treat the disease. Currently, there is debate as to whether lowering or increasing levels of APOE will be beneficial in treating AD. Increasing astrocytic APOE levels can help displace synaptic A β [135] and prevent subsequent synaptotoxicity. Such interventions, with the recent example of Bexarotene, have had mixed outcomes in mouse models of AD [136–139] and no direct benefits so far in human trials [140]. In contrast, lowering levels of APOE4 and increasing levels of APOE2 [59] and APOE3 [141], or decreasing total APOE [142] are promising alternative avenues for maintaining brain resilience and synaptic integrity.

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Author Contributions

MT – design of systematic search, paper screening, writing paper, staining and imaging tissue, revising paper

CD - design of systematic search, paper screening, writing paper, revising paper

AN - design of systematic search, paper screening, writing paper

RJJ – writing paper, revising paper

TS-J – design of systematic search, writing paper, revising paper

Ethics

Use of human tissue for post-mortem studies has been reviewed and approved by the Edinburgh Brain Bank ethics committee and the ACCORD medical research ethics committee, AMREC (approval number 15-HV-016; ACCORD is the Academic and Clinical Central Office for Research and Development, a joint office of the University of Edinburgh and NHS Lothian). The Edinburgh Brain Bank is a Medical Research Council funded facility with research ethics committee (REC) approval (11/ES/0022). Tissue from 4 donors was used for this study and their details are found in supplementary information.

Authors declare no conflicts of interest.

References

- 1 Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, *et al.* Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 1993;**261**:921–3.
- 2 Li YJ, Hauser MA, Scott WK, Martin ER, Booze MW, Qin XJ, *et al.* Apolipoprotein E controls the risk and age at onset of Parkinson disease. *Neurology* 2004;**62**:2005–9.
- 3 Agosta F, Vessel KA, Miller BL, Migliaccio R, Bonasera SJ, Filippi M, *et al.* Apolipoprotein E epsilon4 is associated with disease-specific effects on brain atrophy in Alzheimer's disease and frontotemporal dementia. *Proc Natl Acad Sci USA* 2009;**106**:2018–22.
- 4 Huynh T-PV, Davis AA, Ulrich JD, Holtzman DM. Apolipoprotein E and Alzheimer's disease: the influence of apolipoprotein E on amyloid- β and other amyloidogenic proteins. *J Lipid Res* 2017;**58**:824–36.
- 5 Deary IJ, Whiteman MC, Pattie A, Starr JM, Hayward C, Wright AF, *et al.* Apolipoprotein e gene

variability and cognitive functions at age 79: a follow-up of the Scottish mental survey of 1932. *Psychol Aging* 2004;**19**:367–71.

- 6 Rebeck GW, Reiter JS, Strickland DK, Hyman BT. Apolipoprotein E in sporadic Alzheimer's disease: allelic variation and receptor interactions. *Neuron* 1993;**11**:575–80.
- 7 Liu C-C, Liu C-C, Kanekiyo T, Xu H, Bu G. Apolipoprotein E and Alzheimer disease: risk, mechanisms and therapy. *Nat Rev Neurol* 2013;**9**:106–18.
- 8 De Strooper B, Karran E. The cellular phase of alzheimer's disease. *Cell* 2016;**164**:603–15.
- 9 Shi Y, Yamada K, Liddel SA, Smith ST, Zhao L, Luo W, *et al.* ApoE4 markedly exacerbates tau-mediated neurodegeneration in a mouse model of tauopathy. *Nature* 2017;**549**:523–7.
- 10 Moher D, Liberati A, Tetzlaff J, Altman DG, PRISMA Group. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *PLoS Med* 2009;**6**:e1000097.
- 11 Boccardi M, Sabattoli F, Testa C, Beltramello A, Soininen H, Frisoni GB. APOE and modulation of Alzheimer's and frontotemporal dementia. *Neurosci Lett* 2004;**356**:167–70.
- 12 Filippini N, Rao A, Wetten S, Gibson RA, Borrie M, Guzman D, *et al.* Anatomically-distinct genetic associations of APOE epsilon4 allele load with regional cortical atrophy in Alzheimer's disease. *Neuroimage* 2009;**44**:724–8.
- 13 Gispert JD, Rami L, Sánchez-Benavides G, Falcon C, Tucholka A, Rojas S, *et al.* Nonlinear cerebral atrophy patterns across the Alzheimer's disease continuum: impact of APOE4 genotype. *Neurobiol Aging* 2015;**36**:2687–701.
- 14 Lehtovirta M, Laakso MP, Soininen H, Helisalmi S, Mannermaa A, Helkala EL, *et al.* Volumes of hippocampus, amygdala and frontal lobe in Alzheimer patients with different apolipoprotein E genotypes. *Neuroscience* 1995;**67**:65–72.
- 15 Lehtovirta M, Soininen H, Laakso MP, Partanen K, Helisalmi S, Mannermaa A, *et al.* SPECT and MRI analysis in Alzheimer's disease: relation to apolipoprotein E epsilon 4 allele. *J Neurol Neurosurg Psychiatry* 1996;**60**:644–9.
- 16 Lupton MK, Strike L, Hansell NK, Wen W, Mather KA, Armstrong NJ, *et al.* The effect of increased genetic risk for Alzheimer's disease on hippocampal and amygdala volume. *Neurobiol Aging* 2016;**40**:68–77.
- 17 Geroldi C, Pihlajamäki M, Laakso MP, DeCarli C, Beltramello A, Bianchetti A, *et al.* APOE-epsilon4 is associated with less frontal and more medial temporal lobe atrophy in AD. *Neurology* 1999;**53**:1825–32.
- 18 Juottonen K, Lehtovirta M, Helisalmi S, Riekkinen PJ, Soininen H. Major decrease in the volume of the entorhinal cortex in patients with Alzheimer's disease carrying the apolipoprotein E epsilon4 allele. *J Neurol Neurosurg Psychiatry* 1998;**65**:322–7.
- 19 Hashimoto M, Yasuda M, Tanimukai S, Matsui M, Hirono N, Kazui H, *et al.* Apolipoprotein E epsilon 4 and the pattern of regional brain atrophy in Alzheimer's disease. *Neurology*

2001;**57**:1461–6.

- 20 Wolk DA, Dickerson BC, Alzheimer's Disease Neuroimaging Initiative. Apolipoprotein E (APOE) genotype has dissociable effects on memory and attentional-executive network function in Alzheimer's disease. *Proc Natl Acad Sci USA* 2010;**107**:10256–61.
- 21 Liu Y, Paajanen T, Westman E, Wahlund L-O, Simmons A, Tunnard C, *et al.* Effect of APOE ϵ 4 allele on cortical thicknesses and volumes: the AddNeuroMed study. *J Alzheimers Dis* 2010;**21**:947–66.
- 22 Manning EN, Barnes J, Cash DM, Bartlett JW, Leung KK, Ourselin S, *et al.* APOE ϵ 4 is associated with disproportionate progressive hippocampal atrophy in AD. *PLoS One* 2014;**9**:e97608.
- 23 Susanto TAK, Pua EPK, Zhou J, Alzheimer's Disease Neuroimaging Initiative. Cognition, brain atrophy, and cerebrospinal fluid biomarkers changes from preclinical to dementia stage of Alzheimer's disease and the influence of apolipoprotein e. *J Alzheimers Dis* 2015;**45**:253–68.
- 24 Jack CR, Petersen RC, Xu YC, O'Brien PC, Waring SC, Tangalos EG, *et al.* Hippocampal atrophy and apolipoprotein E genotype are independently associated with Alzheimer's disease. *Ann Neurol* 1998;**43**:303–10.
- 25 Drzezga A, Grimmer T, Henriksen G, Mühlau M, Perneczky R, Miederer I, *et al.* Effect of APOE genotype on amyloid plaque load and gray matter volume in Alzheimer disease. *Neurology* 2009;**72**:1487–94.
- 26 Mori E, Lee K, Yasuda M, Hashimoto M, Kazui H, Hirano N, *et al.* Accelerated hippocampal atrophy in Alzheimer's disease with apolipoprotein E epsilon4 allele. *Ann Neurol* 2002;**51**:209–14.
- 27 Schuff N, Woerner N, Boreta L, Kornfield T, Shaw LM, Trojanowski JQ, *et al.* MRI of hippocampal volume loss in early Alzheimer's disease in relation to ApoE genotype and biomarkers. *Brain* 2009;**132**:1067–77.
- 28 Lo RY, Hubbard AE, Shaw LM, Trojanowski JQ, Petersen RC, Aisen PS, *et al.* Longitudinal change of biomarkers in cognitive decline. *Arch Neurol* 2011;**68**:1257–66.
- 29 Risacher SL, Shen L, West JD, Kim S, McDonald BC, Beckett LA, *et al.* Longitudinal MRI atrophy biomarkers: relationship to conversion in the ADNI cohort. *Neurobiol Aging* 2010;**31**:1401–18.
- 30 Jack CR, Petersen RC, Xu Y, O'Brien PC, Smith GE, Ivnik RJ, *et al.* Rate of medial temporal lobe atrophy in typical aging and Alzheimer's disease. *Neurology* 1998;**51**:993–9.
- 31 Laakso MP, Frisoni GB, Könönen M, Mikkonen M, Beltramello A, Geroldi C, *et al.* Hippocampus and entorhinal cortex in frontotemporal dementia and Alzheimer's disease: a morphometric MRI study. *Biol Psychiatry* 2000;**47**:1056–63.
- 32 Yasuda M, Mori E, Kitagaki H, Yamashita H, Hirano N, Shimada K, *et al.* Apolipoprotein E epsilon 4 allele and whole brain atrophy in late-onset Alzheimer's disease. *Am J Psychiatry* 1998;**155**:779–84.

- 33 Arendt T, Schindler C, Brückner MK, Eschrich K, Bigl V, Zedlick D, *et al.* Plastic neuronal remodeling is impaired in patients with Alzheimer's disease carrying apolipoprotein epsilon 4 allele. *J Neurosci* 1997;**17**:516–29.
- 34 Camicioli R, Kaye J, Payami H, Ball MJ, Murdoch G. Apolipoprotein E epsilon4 is associated with neuronal loss in the substantia nigra in Alzheimer's disease. *Dement Geriatr Cogn Disord* 1999;**10**:437–41.
- 35 Gilmor ML, Erickson JD, Varoqui H, Hersh LB, Bennett DA, Cochran EJ, *et al.* Preservation of nucleus basalis neurons containing choline acetyltransferase and the vesicular acetylcholine transporter in the elderly with mild cognitive impairment and early Alzheimer's disease. *J Comp Neurol* 1999;**411**:693–704.
- 36 Gómez-Isla T, Hollister R, West H, Mui S, Growdon JH, Petersen RC, *et al.* Neuronal loss correlates with but exceeds neurofibrillary tangles in Alzheimer's disease. *Ann Neurol* 1997;**41**:17–24.
- 37 Yin J, Turner G, Coons S, Maalouf M, Reiman E, Shi J. Association of amyloid burden, brain atrophy and memory deficits in aged apolipoprotein ε4 mice. *Curr Alzheimer Res* 2014;**11**:283–90.
- 38 Buttini M, Akeefe H, Lin C, Mahley RW, Pitas RE, Wyss-Coray T, *et al.* Dominant negative effects of apolipoprotein E4 revealed in transgenic models of neurodegenerative disease. *Neuroscience* 2000;**97**:207–10.
- 39 Belinson H, Lev D, Masliah E, Michaelson DM. Activation of the amyloid cascade in apolipoprotein E4 transgenic mice induces lysosomal activation and neurodegeneration resulting in marked cognitive deficits. *J Neurosci* 2008;**28**:4690–701.
- 40 Jordán J, Galindo MF, Miller RJ, Reardon CA, Getz GS, LaDu MJ. Isoform-specific effect of apolipoprotein E on cell survival and beta-amyloid-induced toxicity in rat hippocampal pyramidal neuronal cultures. *J Neurosci* 1998;**18**:195–204.
- 41 Manelli AM, Bulfinch LC, Sullivan PM, LaDu MJ. Abeta42 neurotoxicity in primary co-cultures: effect of apoE isoform and Abeta conformation. *Neurobiol Aging* 2007;**28**:1139–47.
- 42 Drouet B, Fifre A, Pinçon-Raymond M, Vandekerckhove J, Rosseneu M, Guéant JL, *et al.* ApoE protects cortical neurones against neurotoxicity induced by the non-fibrillar C-terminal domain of the amyloid-beta peptide. *J Neurochem* 2001;**76**:117–27.
- 43 Andrews-Zwilling Y, Bien-Ly N, Xu Q, Li G, Bernardo A, Yoon SY, *et al.* Apolipoprotein E4 causes age- and Tau-dependent impairment of GABAergic interneurons, leading to learning and memory deficits in mice. *J Neurosci* 2010;**30**:13707–17.
- 44 Leung L, Andrews-Zwilling Y, Yoon SY, Jain S, Ring K, Dai J, *et al.* Apolipoprotein E4 causes age- and sex-dependent impairments of hilar GABAergic interneurons and learning and memory deficits in mice. *PLoS One* 2012;**7**:e53569.
- 45 Grouselle D, Winsky-Sommerer R, David JP, Delacourte A, Dournaud P, Epelbaum J. Loss of

somatostatin-like immunoreactivity in the frontal cortex of Alzheimer patients carrying the apolipoprotein epsilon 4 allele. *Neurosci Lett* 1998;**255**:21–4.

- 46 Scheff SW, Price DA, Schmitt FA, Mufson EJ. Hippocampal synaptic loss in early Alzheimer's disease and mild cognitive impairment. *Neurobiol Aging* 2006;**27**:1372–84.
- 47 Scheff SW, Price DA, Schmitt FA, DeKosky ST, Mufson EJ. Synaptic alterations in CA1 in mild Alzheimer disease and mild cognitive impairment. *Neurology* 2007;**68**:1501–8.
- 48 Scheff SW, Price DA, Schmitt FA, Scheff MA, Mufson EJ. Synaptic loss in the inferior temporal gyrus in mild cognitive impairment and Alzheimer's disease. *J Alzheimers Dis* 2011;**24**:547–57.
- 49 Cambon K, Davies HA, Stewart MG. Synaptic loss is accompanied by an increase in synaptic area in the dentate gyrus of aged human apolipoprotein E4 transgenic mice. *Neuroscience* 2000;**97**:685–92.
- 50 Heinonen O. Alzheimer pathology of patients carrying apolipoprotein E ϵ 4 allele. *Neurobiol Aging* 1995;**16**:505–13.
- 51 Corey-Bloom J, Tiraboschi P, Hansen LA, Alford M, Schoos B, Sabbagh MN, *et al.* E4 allele dosage does not predict cholinergic activity or synapse loss in Alzheimer's disease. *Neurology* 2000;**54**:403–6.
- 52 Blennow K, Bogdanovic N, Alafuzoff I, Ekman R, Davidsson P. Synaptic pathology in Alzheimer's disease: relation to severity of dementia, but not to senile plaques, neurofibrillary tangles, or the ApoE4 allele. *J Neural Transm* 1996;**103**:603–18.
- 53 Buttini M, Orth M, Bellosta S, Akeefe H, Pitas RE, Wyss-Coray T, *et al.* Expression of human apolipoprotein E3 or E4 in the brains of Apoe^{-/-} mice: isoform-specific effects on neurodegeneration. *J Neurosci* 1999;**19**:4867–80.
- 54 Buttini M, Yu G-Q, Shockley K, Huang Y, Jones B, Masliah E, *et al.* Modulation of Alzheimer-like synaptic and cholinergic deficits in transgenic mice by human apolipoprotein E depends on isoform, aging, and overexpression of amyloid beta peptides but not on plaque formation. *J Neurosci* 2002;**22**:10539–48.
- 55 Veinbergs I, Mallory M, Mante M, Rockenstein E, Gilbert JR, Masliah E. Differential neurotrophic effects of apolipoprotein E in aged transgenic mice. *Neurosci Lett* 1999;**265**:218–22.
- 56 Masliah E, Mallory M, Ge N, Alford M, Veinbergs I, Roses AD. Neurodegeneration in the central nervous system of apoE-deficient mice. *Exp Neurol* 1995;**136**:107–22.
- 57 Anderson R, Barnes JC, Bliss TV, Cain DP, Cambon K, Davies HA, *et al.* Behavioural, physiological and morphological analysis of a line of apolipoprotein E knockout mouse. *Neuroscience* 1998;**85**:93–110.
- 58 Tannenberg RK, Scott HL, Tannenberg AEG, Dodd PR. Selective loss of synaptic proteins in Alzheimer's disease: evidence for an increased severity with APOE varepsilon4. *Neurochem Int*

2006;**49**:631–9.

- 59 Hudry E, Dashkoff J, Roe AD, Takeda S, Koffie RM, Hashimoto T, *et al.* Gene transfer of human Apoe isoforms results in differential modulation of amyloid deposition and neurotoxicity in mouse brain. *Sci Transl Med* 2013;**5**:212ra161.
- 60 Koffie RM, Hashimoto T, Tai H-C, Kay KR, Serrano-Pozo A, Joyner D, *et al.* Apolipoprotein E4 effects in Alzheimer's disease are mediated by synaptotoxic oligomeric amyloid- β . *Brain* 2012;**135**:2155–68.
- 61 Liu D, Pan X, Zhang J, Shen H, Collins NC, Cole AM, *et al.* APOE4 enhances age-dependent decline in cognitive function by down-regulating an NMDA receptor pathway in EFAD-Tg mice. *Mol Neurodegener* 2015;**10**:7.
- 62 Holtzman DM, Bales KR, Tenkova T, Fagan AM, Parsadanian M, Sartorius LJ, *et al.* Apolipoprotein E isoform-dependent amyloid deposition and neuritic degeneration in a mouse model of Alzheimer's disease. *Proc Natl Acad Sci USA* 2000;**97**:2892–7.
- 63 Nwabuisi-Heath E, Rebeck GW, Ladu MJ, Yu C. ApoE4 delays dendritic spine formation during neuron development and accelerates loss of mature spines in vitro. *ASN Neuro* 2014;**6**:e00134.
- 64 Dumanis SB, Tesoriero JA, Babus LW, Nguyen MT, Trotter JH, Ladu MJ, *et al.* ApoE4 decreases spine density and dendritic complexity in cortical neurons in vivo. *J Neurosci* 2009;**29**:15317–22.
- 65 Jain S, Yoon SY, Leung L, Knoferle J, Huang Y. Cellular source-specific effects of apolipoprotein (apo) E4 on dendrite arborization and dendritic spine development. *PLoS One* 2013;**8**:e59478.
- 66 Rodriguez GA, Burns MP, Weeber EJ, Rebeck GW. Young APOE4 targeted replacement mice exhibit poor spatial learning and memory, with reduced dendritic spine density in the medial entorhinal cortex. *Learn Mem* 2013;**20**:256–66.
- 67 Wang C, Wilson WA, Moore SD, Mace BE, Maeda N, Schmechel DE, *et al.* Human apoE4-targeted replacement mice display synaptic deficits in the absence of neuropathology. *Neurobiol Dis* 2005;**18**:390–8.
- 68 Ji Y, Gong Y, Gan W, Beach T, Holtzman DM, Wisniewski T. Apolipoprotein E isoform-specific regulation of dendritic spine morphology in apolipoprotein E transgenic mice and Alzheimer's disease patients. *Neuroscience* 2003;**122**:305–15.
- 69 Lanz TA, Carter DB, Merchant KM. Dendritic spine loss in the hippocampus of young PDAPP and Tg2576 mice and its prevention by the ApoE2 genotype. *Neurobiol Dis* 2003;**13**:246–53.
- 70 Sen A, Alkon DL, Nelson TJ. Apolipoprotein E3 (ApoE3) but not ApoE4 protects against synaptic loss through increased expression of protein kinase C epsilon. *J Biol Chem* 2012;**287**:15947–58.
- 71 LaFerla FM, Troncoso JC, Strickland DK, Kawas CH, Jay G. Neuronal cell death in Alzheimer's disease correlates with apoE uptake and intracellular Abeta stabilization. *J Clin Invest*

1997;**100**:310–20.

- 72 Belinson H, Kariv-Inbal Z, Kayed R, Masliah E, Michaelson DM. Following activation of the amyloid cascade, apolipoprotein E4 drives the in vivo oligomerization of amyloid- β resulting in neurodegeneration. *J Alzheimers Dis* 2010;**22**:959–70.
- 73 Veinbergs I, Everson A, Sagara Y, Masliah E. Neurotoxic effects of apolipoprotein E4 are mediated via dysregulation of calcium homeostasis. *J Neurosci Res* 2002;**67**:379–87.
- 74 Xu Q, Bernardo A, Walker D, Kanegawa T, Mahley RW, Huang Y. Profile and regulation of apolipoprotein E (ApoE) expression in the CNS in mice with targeting of green fluorescent protein gene to the ApoE locus. *J Neurosci* 2006;**26**:4985–94.
- 75 Buttini M, Masliah E, Yu G-Q, Palop JJ, Chang S, Bernardo A, *et al.* Cellular source of apolipoprotein E4 determines neuronal susceptibility to excitotoxic injury in transgenic mice. *Am J Pathol* 2010;**177**:563–9.
- 76 Harris FM, Brecht WJ, Xu Q, Teseur I, Kekonius L, Wyss-Coray T, *et al.* Carboxyl-terminal-truncated apolipoprotein E4 causes Alzheimer's disease-like neurodegeneration and behavioral deficits in transgenic mice. *Proc Natl Acad Sci USA* 2003;**100**:10966–71.
- 77 Chang S, ran Ma T, Miranda RD, Balestra ME, Mahley RW, Huang Y. Lipid- and receptor-binding regions of apolipoprotein E4 fragments act in concert to cause mitochondrial dysfunction and neurotoxicity. *Proc Natl Acad Sci USA* 2005;**102**:18694–9.
- 78 Bien-Ly N, Andrews-Zwilling Y, Xu Q, Bernardo A, Wang C, Huang Y. C-terminal-truncated apolipoprotein (apo) E4 inefficiently clears amyloid-beta (Abeta) and acts in concert with Abeta to elicit neuronal and behavioral deficits in mice. *Proc Natl Acad Sci USA* 2011;**108**:4236–41.
- 79 Dafnis I, Argyri L, Sagnou M, Tzinia A, Tsilibary EC, Stratikos E, *et al.* The ability of apolipoprotein E fragments to promote intraneuronal accumulation of amyloid beta peptide 42 is both isoform and size-specific. *Sci Rep* 2016;**6**:30654.
- 80 Tolar M, Keller JN, Chan S, Mattson MP, Marques MA, Crutcher KA. Truncated apolipoprotein E (ApoE) causes increased intracellular calcium and may mediate ApoE neurotoxicity. *J Neurosci* 1999;**19**:7100–10.
- 81 Zhong N, Searce-Levie K, Ramaswamy G, Weisgraber KH. Apolipoprotein E4 domain interaction: synaptic and cognitive deficits in mice. *Alzheimers Dement* 2008;**4**:179–92.
- 82 Nathan BP, Jiang Y, Wong GK, Shen F, Brewer GJ, Struble RG. Apolipoprotein E4 inhibits, and apolipoprotein E3 promotes neurite outgrowth in cultured adult mouse cortical neurons through the low-density lipoprotein receptor-related protein. *Brain Res* 2002;**928**:96–105.
- 83 Korwek KM, Trotter JH, Ladu MJ, Sullivan PM, Weeber EJ. ApoE isoform-dependent changes in hippocampal synaptic function. *Mol Neurodegener* 2009;**4**:21.
- 84 Neustadt AL, Winston CN, Parsadanian M, Main BS, Villapol S, Burns MP. Reduced cortical

excitatory synapse number in APOE4 mice is associated with increased calcineurin activity. *Neuroreport* 2017;**28**:618–24.

- 85 Minett T, Classey J, Matthews FE, Fahrenhold M, Taga M, Brayne C, *et al.* Microglial immunophenotype in dementia with Alzheimer's pathology. *J Neuroinflammation* 2016;**13**:135.
- 86 Egensperger R, Kösel S, von Eitzen U, Graeber MB. Microglial activation in Alzheimer disease: Association with APOE genotype. *Brain Pathol* 1998;**8**:439–47.
- 87 Overmyer M, Helisalmi S, Soininen H, Laakso M, Riekkinen P, Alafuzoff I. Astrogliosis and the ApoE genotype. an immunohistochemical study of postmortem human brain tissue. *Dement Geriatr Cogn Disord* 1999;**10**:252–7.
- 88 Rodriguez GA, Tai LM, LaDu MJ, Rebeck GW. Human APOE4 increases microglia reactivity at A β plaques in a mouse model of A β deposition. *J Neuroinflammation* 2014;**11**:111.
- 89 Liu C-C, Zhao N, Fu Y, Wang N, Linares C, Tsai C-W, *et al.* Apoe4 accelerates early seeding of amyloid pathology. *Neuron* 2017;**96**:1024–1032.e3.
- 90 Belinson H, Michaelson DM. ApoE4-dependent Abeta-mediated neurodegeneration is associated with inflammatory activation in the hippocampus but not the septum. *J Neural Transm* 2009;**116**:1427–34.
- 91 Ophir G, Meilin S, Efrati M, Chapman J, Karussis D, Roses A, *et al.* Human apoE3 but not apoE4 rescues impaired astrocyte activation in apoE null mice. *Neurobiol Dis* 2003;**12**:56–64.
- 92 Kang SS, Ebbert MTW, Baker KE, Cook C, Wang X, Sens JP, *et al.* Microglial translational profiling reveals a convergent APOE pathway from aging, amyloid, and tau. *J Exp Med* 2018;**215**:2235–45.
- 93 Krasemann S, Madore C, Cialic R, Baufeld C, Calcagno N, El Fatimy R, *et al.* The TREM2-APOE Pathway Drives the Transcriptional Phenotype of Dysfunctional Microglia in Neurodegenerative Diseases. *Immunity* 2017;**47**:566–581.e9.
- 94 Keren-Shaul H, Spinrad A, Weiner A, Matcovitch-Natan O, Dvir-Szternfeld R, Ulland TK, *et al.* A Unique Microglia Type Associated with Restricting Development of Alzheimer's Disease. *Cell* 2017;**169**:1276–1290.e17.
- 95 Lin Y-T, Seo J, Gao F, Feldman HM, Wen H-L, Penney J, *et al.* APOE4 Causes Widespread Molecular and Cellular Alterations Associated with Alzheimer's Disease Phenotypes in Human iPSC-Derived Brain Cell Types. *Neuron* 2018;**98**:1141–1154.e7.
- 96 Caberlotto L, Marchetti L, Lauria M, Scotti M, Parolo S. Integration of transcriptomic and genomic data suggests candidate mechanisms for APOE4-mediated pathogenic action in Alzheimer's disease. *Sci Rep* 2016;**6**:32583.
- 97 Thangavel R, Bhagavan SM, Ramaswamy SB, Surpur S, Govindarajan R, Kempuraj D, *et al.* Co-Expression of Glia Maturation Factor and Apolipoprotein E4 in Alzheimer's Disease Brain. *J*

Alzheimers Dis 2018;**61**:553–60.

- 98 Vitek MP, Brown CM, Colton CA. APOE genotype-specific differences in the innate immune response. *Neurobiol Aging* 2009;**30**:1350–60.
- 99 Tulloch J, Leong L, Thomson Z, Chen S, Lee E-G, Keene CD, *et al.* Glia-specific APOE epigenetic changes in the Alzheimer's disease brain. *Brain Res* Published Online First: 3 August 2018. doi:10.1016/j.brainres.2018.08.006
- 100 Colton CA, Brown CM, Cook D, Needham LK, Xu Q, Czapiga M, *et al.* APOE and the regulation of microglial nitric oxide production: a link between genetic risk and oxidative stress. *Neurobiol Aging* 2002;**23**:777–85.
- 101 Brown CM, Wright E, Colton CA, Sullivan PM, Laskowitz DT, Vitek MP. Apolipoprotein E isoform mediated regulation of nitric oxide release. *Free Radic Biol Med* 2002;**32**:1071–5.
- 102 Chen S, Averett NT, Manelli A, LaDu MJ, May W, Ard MD. Isoform-specific effects of apolipoprotein E on secretion of inflammatory mediators in adult rat microglia. *JAD* 2005;**7**:25–35.
- 103 Guo L, LaDu MJ, Van Eldik LJ. A dual role for apolipoprotein e in neuroinflammation: anti- and pro-inflammatory activity. *J Mol Neurosci* 2004;**23**:205–12.
- 104 Hu J, LaDu MJ, Van Eldik LJ. Apolipoprotein E attenuates beta-amyloid-induced astrocyte activation. *J Neurochem* 1998;**71**:1626–34.
- 105 Dorey E, Bamji-Mirza M, Najem D, Li Y, Liu H, Callaghan D, *et al.* Apolipoprotein E Isoforms Differentially Regulate Alzheimer's Disease and Amyloid- β -Induced Inflammatory Response in vivo and in vitro. *J Alzheimers Dis* 2017;**57**:1265–79.
- 106 Maezawa I, Nivison M, Montine KS, Maeda N, Montine TJ. Neurotoxicity from innate immune response is greatest with targeted replacement of E4 allele of apolipoprotein E gene and is mediated by microglial p38MAPK. *FASEB J* 2006;**20**:797–9.
- 107 Tai LM, Ghura S, Koster KP, Liakaite V, Maienschein-Cline M, Kanabar P, *et al.* APOE-modulated A β -induced neuroinflammation in Alzheimer's disease: current landscape, novel data, and future perspective. *J Neurochem* 2015;**133**:465–88.
- 108 Laskowitz DT, Thekdi AD, Thekdi SD, Han SK, Myers JK, Pizzo SV, *et al.* Downregulation of microglial activation by apolipoprotein E and apoE-mimetic peptides. *Exp Neurol* 2001;**167**:74–85.
- 109 Ulrich JD, Ulland TK, Mahan TE, Nyström S, Nilsson KP, Song WM, *et al.* ApoE facilitates the microglial response to amyloid plaque pathology. *J Exp Med* 2018;**215**:1047–58.
- 110 Laskowitz DT, Matthew WD, Bennett ER, Schmechel D, Herbstreith MH, Goel S, *et al.* Endogenous apolipoprotein E suppresses LPS-stimulated microglial nitric oxide production. *Neuroreport* 1998;**9**:615–8.
- 111 Lynch JR, Morgan D, Mance J, Matthew WD, Laskowitz DT. Apolipoprotein E modulates glial

activation and the endogenous central nervous system inflammatory response. *J Neuroimmunol* 2001;**114**:107–13.

- 112 Maezawa I, Maeda N, Montine TJ, Montine KS. Apolipoprotein E-specific innate immune response in astrocytes from targeted replacement mice. *J Neuroinflammation* 2006;**3**:10.
- 113 Ophir G, Amariglio N, Jacob-Hirsch J, Elkon R, Rechavi G, Michaelson DM. Apolipoprotein E4 enhances brain inflammation by modulation of the NF-kappaB signaling cascade. *Neurobiol Dis* 2005;**20**:709–18.
- 114 Blain J-F, Sullivan PM, Poirier J. A deficit in astroglial organization causes the impaired reactive sprouting in human apolipoprotein E4 targeted replacement mice. *Neurobiol Dis* 2006;**21**:505–14.
- 115 Cudaback E, Li X, Montine KS, Montine TJ, Keene CD. Apolipoprotein E isoform-dependent microglia migration. *FASEB J* 2011;**25**:2082–91.
- 116 Cudaback E, Yang Y, Montine TJ, Keene CD. APOE genotype-dependent modulation of astrocyte chemokine CCL3 production. *Glia* 2015;**63**:51–65.
- 117 Gomez-Arboledas A, Davila JC, Sanchez-Mejias E, Navarro V, Nuñez-Diaz C, Sanchez-Varo R, *et al.* Phagocytic clearance of presynaptic dystrophies by reactive astrocytes in Alzheimer's disease. *Glia* 2018;**66**:637–53.
- 118 Hong S, Beja-Glasser VF, Nfonoyim BM, Frouin A, Li S, Ramakrishnan S, *et al.* Complement and microglia mediate early synapse loss in Alzheimer mouse models. *Science* 2016;**352**:712–6.
- 119 Zhu Y, Nwabuisi-Heath E, Dumanis SB, Tai LM, Yu C, Rebeck GW, *et al.* APOE genotype alters glial activation and loss of synaptic markers in mice. *Glia* 2012;**60**:559–69.
- 120 McGeer PL, Walker DG, Pitas RE, Mahley RW, McGeer EG. Apolipoprotein E4 (ApoE4) but not ApoE3 or ApoE2 potentiates β -amyloid protein activation of complement in vitro. *Brain Res* 1997;**749**:135–8.
- 121 Chung W-S, Verghese PB, Chakraborty C, Joung J, Hyman BT, Ulrich JD, *et al.* Novel allele-dependent role for APOE in controlling the rate of synapse pruning by astrocytes. *Proc Natl Acad Sci USA* 2016;**113**:10186–91.
- 122 Filipello F, Morini R, Corradini I, Zerbi V, Canzi A, Michalski B, *et al.* The microglial innate immune receptor TREM2 is required for synapse elimination and normal brain connectivity. *Immunity* 2018;**48**:979–991.e8.
- 123 Vainchtein ID, Chin G, Cho FS, Kelley KW, Miller JG, Chien EC, *et al.* Astrocyte-derived interleukin-33 promotes microglial synapse engulfment and neural circuit development. *Science* 2018;**359**:1269–73.
- 124 Liddel SA, Guttenplan KA, Clarke LE, Bennett FC, Bohlen CJ, Schirmer L, *et al.* Neurotoxic reactive astrocytes are induced by activated microglia. *Nature* 2017;**541**:481–7.
- 125 Azevedo EP, Ledo JH, Barbosa G, Sobrinho M, Diniz L, Fonseca ACC, *et al.* Activated microglia

mediate synapse loss and short-term memory deficits in a mouse model of transthyretin-related oculoleptomeningeal amyloidosis. *Cell Death Dis* 2013;**4**:e789.

- 126 Braak H, Braak E. Neuropathological stageing of Alzheimer-related changes. *Acta Neuropathol* 1991;**82**:239–59.
- 127 Victoria GS, Zurzolo C. The spread of prion-like proteins by lysosomes and tunneling nanotubes: Implications for neurodegenerative diseases. *J Cell Biol* 2017;**216**:2633–44.
- 128 Pickett EK, Henstridge CM, Allison E, Pitstick R, Pooler A, Wegmann S, *et al.* Spread of tau down neural circuits precedes synapse and neuronal loss in the rTgTauEC mouse model of early Alzheimer's disease. *Synapse* 2017;**71**. doi:10.1002/syn.21965
- 129 Maphis N, Xu G, Kokiko-Cochran ON, Jiang S, Cardona A, Ransohoff RM, *et al.* Reactive microglia drive tau pathology and contribute to the spreading of pathological tau in the brain. *Brain* 2015;**138**:1738–55.
- 130 Prada I, Gabrielli M, Turola E, Iorio A, D'Arrigo G, Parolisi R, *et al.* Glia-to-neuron transfer of miRNAs via extracellular vesicles: a new mechanism underlying inflammation-induced synaptic alterations. *Acta Neuropathol* 2018;**135**:529–50.
- 131 Asai H, Ikezu S, Tsunoda S, Medalla M, Luebke J, Haydar T, *et al.* Depletion of microglia and inhibition of exosome synthesis halt tau propagation. *Nat Neurosci* 2015;**18**:1584–93.
- 132 Rostami J, Holmqvist S, Lindström V, Sigvardson J, Westermark GT, Ingelsson M, *et al.* Human Astrocytes Transfer Aggregated Alpha-Synuclein via Tunneling Nanotubes. *J Neurosci* 2017;**37**:11835–53.
- 133 DeKosky ST, Scheff SW. Synapse loss in frontal cortex biopsies in Alzheimer's disease: correlation with cognitive severity. *Ann Neurol* 1990;**27**:457–64.
- 134 Munafò MR, Davey Smith G. Robust research needs many lines of evidence. *Nature* 2018;**553**:399–401.
- 135 Lane-Donovan C, Herz J. Apoe, apoe receptors, and the synapse in alzheimer's disease. *Trends Endocrinol Metab* 2017;**28**:273–84.
- 136 O'Hare E, Jeggo R, Kim E-M, Barbour B, Walczak J-S, Palmer P, *et al.* Lack of support for bexarotene as a treatment for Alzheimer's disease. *Neuropharmacology* 2016;**100**:124–30.
- 137 LaClair KD, Manaye KF, Lee DL, Allard JS, Savonenko AV, Troncoso JC, *et al.* Treatment with bexarotene, a compound that increases apolipoprotein-E, provides no cognitive benefit in mutant APP/PS1 mice. *Mol Neurodegener* 2013;**8**:18.
- 138 Cramer PE, Cirrito JR, Wesson DW, Lee CYD, Karlo JC, Zinn AE, *et al.* ApoE-directed therapeutics rapidly clear β -amyloid and reverse deficits in AD mouse models. *Science* 2012;**335**:1503–6.
- 139 Tesseur I, Lo AC, Roberfroid A, Dietvorst S, Van Broeck B, Borgers M, *et al.* Comment on "ApoE-directed therapeutics rapidly clear β -amyloid and reverse deficits in AD mouse

models". *Science* 2013;**340**:924–e.

- 140 Cummings JL, Zhong K, Kinney JW, Heaney C, Moll-Tudla J, Joshi A, *et al.* Double-blind, placebo-controlled, proof-of-concept trial of bexarotene in moderate Alzheimer's disease. *Alzheimers Res Ther* 2016;**8**:4.
- 141 McColl BW, McGregor AL, Wong A, Harris JD, Amalfitano A, Magnoni S, *et al.* APOE epsilon3 gene transfer attenuates brain damage after experimental stroke. *J Cereb Blood Flow Metab* 2007;**27**:477–87.
- 142 Huynh T-PV, Liao F, Francis CM, Robinson GO, Serrano JR, Jiang H, *et al.* Age-Dependent Effects of apoE Reduction Using Antisense Oligonucleotides in a Model of β -amyloidosis. *Neuron* 2017;**96**:1013–1023.e4.

Figure Legends

Figure 1. PRISMA flow diagram. Summary of the review process. Template edited from [10]. Papers referenced in the main body of text were identified through our systematic search and can be found in Supplemental Table 3.

Figure 2. Amyloid- β pathology in APOE3/3 and APOE3/4 carriers in normal ageing and AD. A β plaque deposition is evident in aged controls (A) and is exacerbated in the presence of the APOE4 allele (B), resembling an AD-like phenotype. Both APOE3/3 and APOE3/4 AD cases (C and D, respectively) have substantial A β deposition in all six layers of the cortex. Images taken from the grey matter of inferior temporal lobe (Brodmann area [BA] 20/21). Information about all participants donated tissue can be found in Supplemental Table 4. A β is stained with 6F/3D (mouse monoclonal, DAKO, M087201-2, 1:100, 98% formic acid, 5 minutes). Scale bar 1mm.

Figure 3. Tau pathology in APOE3/3 and APOE3/4 carriers in normal ageing and AD. In aged controls, phosphorylated tau species are absent in APOE 3/3 cases (A) and rarely found in APOE3/4 cases (B). In AD, both APOE3/3 (C) and APOE3/4 (D) have markedly increased numbers of tau-positive neurons. Images taken from the grey matter of inferior temporal lobe (BA20/21). Phosphorylated tau is stained with AT8 (mouse monoclonal, ThermoFisher, 1020, 1:2500). Scale bar 50 μ m, insert scale bar 25 μ m.

Figure 4. Activated microglia (CD68) in APOE3/3 and APOE3/4 carriers in normal ageing and AD. The lysosomal marker of microglia and macrophages, CD68, shown in AD (C-D) and age-matched control cases (A-B). Various microglial morphologies can be observed, for example ramified (A and

D) and amoeboid (B and C) in both ageing/AD and APOE3/x. Images taken from the grey matter of inferior temporal lobe (BA20/21). Microglia are stained with CD68 (mouse monoclonal, DAKO, M0876, 1:100, citric acid antigen retrieval). Scale bar 100µm, insert scale bar 25µm.

Figure 5. Activated astrocytes in APOE3/3 and APOE3/4 carriers in normal ageing and AD. Glial fibrillary acid protein (GFAP) is a cytoskeletal protein in activated astrocytes. Activated astrocytes are seen in both ageing (A-B) and AD (C-D), but more pronounced astrogliosis is observed in AD. In AD, astrocytes express more GFAP in the cell bodies, thus appearing darker, especially in APOE3/4 cases (D), and have more processes (C-D). Images taken from the grey matter of inferior temporal lobe (BA20/21). Astrocytes are stained with GFAP (rabbit polyclonal, DAKO, 0334, 1:800). Scale bar 200µm, insert scale bar 100µm.

Figure 6. APOE at the interface of inflammation and neurodegeneration via glial-mediated mechanisms. Microglia and astrocytes expressing APOE4 promote parenchymal gliosis and release pro-inflammatory signals that are potentially associated with synaptic and neuronal loss. The paracrine signalling of microglial mediators along with the APOE-TREM2 pathway induces a pro-inflammatory phenotype, creating a vicious-cycle of inflammation and neurodegeneration. Ineffective clearance of excess synapses by astrocytes in APOE4 mice allows accumulating levels of C1q that can act as a tag for synaptic elimination later in life.

Figure 7. Schematic diagram of pathological protein spread. While direct evidence for APOE influencing the spread of tau through the brain is lacking, the papers in our systematic literature search implicate APOE in many processes that could influence spread. These include synaptic transfer through the synaptic cleft (1), via nanotubes (2), by glial phagocytosis (3), or vesicular secretion (4 & 5).













